

COURSE LAYOUT

1. GENERAL

SCHOOL	APPLIED BIOLOGY & BIOTECHNOLOGY		
DEPARTMENT	BIOTECHNOLOGY		
STUDY LEVEL	<i>Undergraduate</i>		
COURSE CODE	218	SEMESTER	6th
COURSE TITLE	BIOPHYSICS		
INDEPENDENT TEACHING ACTIVITIES		WEEKLY TEACHING HOURS	ECTS
LECTURES		3	3
PRACTICAL EXERCISES		2	2
TOTAL			5
COURSE TYPE	Scientific Specialization		
PREREQUISITES	Physics, Biochemistry		
LANGUAGE	Greek with English support in terminology		
IS THE COURSE OFFERED for ERASMUS STUDENTS?	YES (in English)		
COURSE WEB PAGE	http://openeclass.aua.gr/courses/BIOTECH131/		

2. LEARNING OUTCOMES

Learning Outcomes

The course is a basic introductory course in Biophysics techniques used in the analysis of the structure of biomolecules (proteins, DNA, RNA) such as X-ray crystallography, scattering techniques, multidimensional NMR, molecular dynamics as well as other quantitative techniques such as calorimetry, circular dichroism and fluorescence spectroscopy. Finally, the course aims to help students understand the applications of these techniques in the design of drugs and other bioactive molecules.

Upon successful completion of this course the student will be able to

- Have an understanding the basic features of the biophysical methods
- Is capable of knowing when to use these methodologies
- Analyze and calculate basic information
- Present the results of a relevant study

General Competences

Search , analyze and synthesize data and information, and the use of essential technologies
Teamwork

Work in a multidisciplinary environment

Search , analyze and synthesize data and information, and the use of essential technologies

3. COURSE CONTENT

Theory: Review of the structure of biological macromolecules. X-ray Diffraction. Crystals. Crystallization. Theory of x-ray diffraction. Reciprocal space. Crystallographic symmetry. Structure factors and Intensities. Data Collection. Electron Density Function. Approaches to the Phase Problem. Structure refinement. Radiation scattering from solutions of macromolecules. Thermodynamics and Biochemistry. Calorimetric methods. Molecular Mechanics, NMR Spectroscopy, circular dichroism, fluorescence spectroscopy, Applications in drug design and Nanotechnology

Laboratory: Determination of thermodynamic parameters for salt dissolution, crystallization of lysozyme, diffraction experiments with lysozyme crystals, analysis of electron density map for lysozyme-ligand complex, fluorescence microscopy image processing.

4. TEACHING and LEARNING METHODS - Evaluation

TEACHING METHOD	In suitably equipped teaching rooms	
USE OF INFORMATICS and COMMUNICATION TECHNOLOGIES	Use of powerpoint presentations and Phet/e-crystallography simulations in lectures, use of specialized software such as the WinCoot, use of e-class website and videos to inform, educate and communicate with students	
TEACHING ORGANISATION	<i>Activity</i>	<i>Work Load</i>
	Lectures	39
	Laboratory exercises	20
	Educational visit	10
	Independent study	56
	<i>Course total (25 hours of student work load per ECTS)</i>	125
STUDENTS EVALUATION	<p>I. Theory: Written final examination (100%) comprising: multiple choice questions, problem solving and short answer questions. Optional exercises during lectures (bonus 10% grade)</p> <p>II. Laboratory: Tests before each laboratory session (15%), written assignments on the laboratory exercises (50%), written final examination (35%).</p>	

5. BIBLIOGRAPHY

1. Principles in Physical Biochemistry (van Holde, Johnson, Ho) 2nd Edition
2. Themata Moriakis Biofysikis (Hamodrakas) Symmetria publications