COURSE OUTLINE

1. GENERAL

SCHOOL	APPLIED BIOLOGY AND BIOTECHNOLOGY					
ACADEMIC UNIT	BIOTECHNOLOGY					
LEVEL OF STUDIES	Undergraduate					
COURSE CODE	175	SEMESTER 7th				
COURSE TITLE	ANIMAL BIOTECHNOLOGY					
INDEPENDENT TEACHII	NG ACTIVITIES WEEKLY TEACHING CREDITE HOURS			DITS		
LECTURES			3	0,	12	
PRACTICAL EXERCISES		2	0,	80		
ESSAYS		1	0,	04		
TOTAL ECTS (Table 4)			5,	00		
COURSE TYPE general background, special background, specialised general knowledge, skills development	Specialised g	eneral knowledg	ge, Skills develo	pment		
PREREQUISITE COURSES:	No					
LANGUAGE OF INSTRUCTION and EXAMINATIONS:	Greek					
IS THE COURSE OFFERED TO ERASMUS STUDENTS	Yes (in English)					
COURSE WEBSITE (URL)	https://mediasrv.aua.gr/eclass/courses/BIOTECH142/					

2. LEARNING OUTCOMES

LEARNING OUTCOMES

The course learning outcomes, specific knowledge, skills and competences of an appropriate level, which the students will acquire with the successful completion of the course are described.

Consult Appendix A

- Description of the level of learning outcomes for each qualifications cycle, according to the Qualifications
 Framework of the European Higher Education Area
- Descriptors for Levels 6, 7 & 8 of the European Qualifications Framework for Lifelong Learning and Appendix B
- Guidelines for writing Learning Outcomes

The course is a basic introductory course on concepts of animal biotechnology, genetic engineering technologies in animal organisms and the applications arising in the agricultural and biomedical field.

This course aims to introduce students to the basic technologies of animal biotechnology for the understanding and application of genetic engineering technologies related to the creation of transgenic animals, gene targeting for gene inactivation, modification and conditional gene expression, as well as to the identification of mutations by genetic analysis. It also refers to basic concepts and methodologies for the generation of biotechnological products such as monoclonal antibodies, recombinant proteins, vaccines and latest technologies related to animal cloning, stem cell biology, regenerative medicine, gene therapy, animal cell culturing, cryopreservation, assisted reproduction, genotypic analysis and diagnosis of diseases in order for the student to obtain a comprehensive insight into the applications of Biotechnology in Animal husbandry and biomedicine.

Finally, the aim of the course is the understanding of the importance of biotechnological technologies based on the use of animal organisms for the identification of gene function and the development of innovative approaches and biotechnological products with applications in diagnosis and treatment.

Upon successful completion of this course the student will be able to:

- Understand the basic approaches of animal biotechnology and its applications in the agricultural and biomedical field.
- Acquire knowledge of the tools and techniques of genetic engineering in animal organisms and their application for the understanding of gene function and the production of innovative biotechnological processes and products.
- Assess the applications of animal biotechnology technologies and to propose them in specific projects / programs.
- Be trained in biotechnological approaches for the production of pharmaceutical proteins, and in modern techniques used for the production of transgenic animals.
- Learn in detail about the process of creating transgenic animals through the method of
 microinjection of DNA in pronuclei of fertilized oocytes but also with lentiviruses, having
 acquired knowledge about the applications of transgenic animals.
- Understand the stages of gene targeting to create knockout mice and its applications.
- Understand the mechanism of action and usefulness of the nucleases ZFNs in genome modifications.
- Understand why the CRISPR / CAS9 system is superior as a genetic tool for modifying animal organisms compared to other techniques such as genetic targeting, ZNFs and TALENs nucleases.
- Distinguish and select genetic tools for the application of gene modifications in mammals either in specific cell types or at specific time intervals.
- Understand the principle and the usefulness of Forward Genetics in the identification of genes involved in the pathogenesis of genetic diseases.
- Acquire knowledge of mammalian cloning methods with an emphasis on nuclear transfer.
- Become familiar with technologies for the production of monoclonal antibodies and with their application in the diagnosis and treatment of human diseases.
- Understand the methods for the production of vaccines and the importance of vaccination to protect population health.
- Understand the importance of the stem cells in combination with gene therapy for the treatment of human diseases.
- Gain full knowledge of cutting-edge technologies based on the use of animal organisms and their cells in order to indulge further by applying them within postgraduate and professional level.
- In the laboratory, the student is trained in tools and cutting-edge techniques of Animal Biotechnology, and collaborates with his fellow students to evaluate the experimental results and adequately present the results and conclusions of the experimental process.

General Competences

Taking into consideration the general competences that the degree-holder must acquire (as these appear in the Diploma Supplement and appear below), at which of the following does the course aim?

Search for, analysis and synthesis of data and information, with the use of the necessary technology

information, with the use of the necessary technology Adapting to new situations

Decision-making
Working independently
Team work

Working in an international environment Working in an interdisciplinary environment Production of new research ideas Project planning and management Respect for differences and multiculturalism Respect for the natural environment

Showing social, professional and ethical responsibility and sensitivity to gender issues

Criticism and self-criticism

Production of free, creative and inductive thinking

- Working independently
- Team work

- Decision-making
- Working in an interdisciplinary environment
- Production of new research ideas
- Project planning and management
- Search for, analysis and synthesis of data and information, with the use of the necessary technology
- · Production of free, creative and inductive thinking

3. SYLLABUS

- 1. <u>Introduction to Animal Biotechnology</u>: Production of pharmaceutical proteins with biotechnological methods. Transgenic technologies in animal organisms. The laboratory mouse as a model organism. Biology, genetics and genome features of the laboratory mouse. Oogenesis, spermatogenesis and fertilization in mammals. Critical embryo stages in Animal Biotechnology. Bioethical issues.
- **2.** <u>Technologies for the generation of transgenic animals:</u> Functional Genomics Technologies. Forward and Reverse Genetics. Creation of transgenic animals through DNA microinjection into the pronuclei of fertilized oocytes. Creating transgenic animals with lentiviruses. Advantages, Disadvantages. Transgenic animal applications.
- **3.** <u>Gene targeting technology</u>: Embryonic stem cells. Design of gene construct. Positive and negative selection markers. Gene targeting stages for the generation of knockout mice. Applications.
- **4.** Genome modifications of animals with zinc-finger nucleases (ZFNs): ZFNs features and mechanism of action. Target Recognition by ZFNs. Design and construction of ZFNs. Mechanisms of genomic modifications with ZFNs. Use in animal organisms and applications.
- **5.** Genome modifications of animals with the system CRISPR/CAS9: The CRISPR/CAS gene locus in the streptococcus. Description of the CRISPR/CAS9 system. Biotechnological use of the CRISPR/CAS9 system in animal organisms. Strategies to minimize off-target modifications. Comparison with ZNFs and TALENs nucleases.
- 6. Spatial and temporal control of inducible gene expression and modifications in animals: Recombination with the Cre/loxP system. Applications of the Cre/loxP system to animal organisms. Tissue-specific gene activation and inactivation. Inducible temporal gene modifications at the transcriptional and post-transcriptional level.
- 7. Forward Genetics: from the phenotype to the identification of the causal gene: Chemical mutagenesis with N-ethyl-N-nitrosourea. Phenotypic analysis. Detection of the mutated gene by mapping. Stages of mapping. Polymorphic genetic markers.
- **8.** <u>Animal cloning</u>: Animal cloning methods. Nuclear transfer. Cloning stages. Reprogramming in cloning. Applications of animal cloning in agriculture and biotechnology. Bioethical issues.
- **9.** <u>Stem cells</u>: Types of mammalian stem cells and their topology. Embryonic stem cells. Adult stem cells. Induced pluripotent stem cells. Stem cell applications.
- **10.** Gene therapy: History of gene therapy. Disease targets. Prerequisites and tools in gene therapy. Ex vivo, in vivo gene therapy applications. Vectors in gene therapy. Stages of gene therapy. Problems in gene therapy. Latest achievements.
- **11.**<u>Monoclonal antibodies Vaccines</u>: Review of the immune system. Monoclonal, polyclonal antibodies. Techniques for the production of monoclonal antibodies. Applications of monoclonal antibodies in the diagnosis and treatment of human diseases. Types of vaccines. Vaccination mechanisms. Examples, HPV vaccines.

<u>Laboratory</u>: Generation of transgenic mice - Design of transgene construct - Isolation of genomic DNA from mouse tissue - Genotyping of transgenic mice - Animal cell culture - Overproduction of proteins in mammalian cell lines - Cryopreservation - *In vitro* fertilization

(IVF), and intracytoplasmic sperm injection (ICSI) - Design of gene inactivation with the CRISPR / CAS9 system.

4. TEACHING and LEARNING METHODS - EVALUATION

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DELIVERY	race to tace in cantaci, equipped teaching recine				
Face-to-face, Distance learning, etc.	Distant Learning.				
USE OF INFORMATION AND	The course is completely computerised in the form of				
COMMUNICATIONS TECHNOLOGY Use of ICT in teaching, laboratory education,	Powerpoint, Web linking, etc.				
communication with students	Computer programmes and applications are taught				
	and distributed to students.				
	Course material also made available to the students via				
	the e-class platform.				
TEACHING METHODS	Activity	Semester Workload			
The manner and methods of teaching are described in detail.	Lectures	39 h			
Lectures, seminars, laboratory practice,	Laboratory work	26 h			
fieldwork, study and analysis of bibliography, tutorials, placements, clinical practice, art	Group and/or individual	13 h			
workshop, interactive teaching, educational	works				
visits, project, essay writing, artistic creativity,	Independent study	47 h			
etc.	Course total (Total				
The student's study hours for each learning	contact hours and	125 h			
activity are given as well as the hours of non- directed study according to the principles of the	training)				
ECTS					
STUDENT PERFORMANCE					
EVALUATION	I. Written final examination in Theory (50%) of				
Description of the evaluation procedure	different difficulty, based on the lectures offered				
Language of evaluation, methods of	of containing:				
evaluation, summative or conclusive, multiple choice questionnaires, short-answer questions,	- Questions of multiple choi	re .			
open-ended questions, problem solving, written	·				
work, essay/report, oral examination, public presentation, laboratory work, clinical	- Questions of theoretical knowledge.				
examination of patient, art interpretation, other	- Problems based on lecture material.				
Charifically defined applications within	II. Laboratory exercises/pra	acticals: Written			
Specifically-defined evaluation criteria are given, and if and where they are accessible to	Examination (35%) of differ				
students.	lectures offered, containing:				
	- Questions of multiple choice.				
	·				
	- Questions of theoretical knowledge.				
	- Problems based on lecture material.				

5. ATTACHED BIBLIOGRAPHY

-Suggested bibliography :

-Relevant scientific journals:

- Molecular Biotechnology: Principles and Applications of Recombinant DNA. Glick BR, Pasternak JJ, Patten CL. 4th edition, published by ASM Press. 2009.
- Principles of Gene Manipulation and Genomics. Twyman R, Primrose SB. 8th edition, Wiley-Blackwell, 2013.

III. Group and small autonomous works (15%).